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Genetic Identification and Phylogeny of *Clerodendrum Phlomidis* Growing in Egypt Using Some DNA Barcodes

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ABSTRACT

Clerodendrum is one of the Lamiaceae genera, which has significance in taxonomical position. Clerodendrum phlomidis is an important wild species in Egypt and plays a main role in ecological, biodiversity, and taxonomical approaches. In the current study, we conducted this research work to investigate the identification of C. phlomidis collected from the North Coastal region of Egypt using three DNA barcode sequences viz., rbcL, ITS, and rpoC1 genes. The investigation was conducted in the corporation between the Agric. Botany-Genetics Dept., Fac. of Agric, Al-Azhar Univ. and National Gene Bank (NGB), Agricultural Research Center (ARC), Egypt. Our findings revealed that accessions KF199889 (98.7%) and ON010669.1 (98.6%) were identical to the taxon of C. phlomidis by the ITS and rbcL sequences. On the other hand, the sequence of C. phlomidis revealed closer to the taxa of C. buchneri, C. cephalanthum, C. thomsoniae, and C. rotundifoliuma by ITS sequences, C. indicum, C. thomsoniae, C. longiflorum, C. infortunatum, C. floribundum, C. cephalanthum, C. buchneri, and C. splendens by rbcL, and C. japonicum, C. thomsoniae, C. cyrtophyllum, and C. trichotomum by rpoC1 sequences. It can be recommended that these partial sequences are accurate genes for identifying and verifying the species of *Clerodendrum phlomidis*.

KEYWORDS: Clerodendrum phlomidis, ITS, rbcL, rpoC1, Molecular Identification, DNA barcoding, BLAST

1. INTRODUCTION

The Lamiaceae family, commonly known as the mint family, is a diverse and economically

significant plant family with a global distribution. With over 200 genera and around 7,000 species, this family includes many well-known herbs and ornamental plants, contributing to various aspects of human life. Lamiaceae family is globally significant due to its economic, culinary, medicinal, and ecological roles. Its members play a vital part in human culture and are integral to various industries, making the family a subject of interest for researchers, horticulturists, and conservationists worldwide (Carović-Stanko et al., 2016). Clerodendrum (Lamiaceae) has about 580 species distributed over the world. Some species viz., C. indicum, C. phlomidis, C. serratum, and C. trichotomum used for the treatment of various diseases. Clerodendrum phlomidis (syn. Volkameria multiflorum) is an erect or scrambling shrub and is commonly found in Egypt. This taxon has significance in ecological, taxonomical, and ethnobotanical approaches, as well as its use as a remedy for various diseases such as diabetes, hypertension, headaches, wounds, snakebites and toothaches. C. phlomidis identification is needed to reduce substitution and the possibility of contamination. (Adamu et al., 2020)(Newmaster et al., 2013). Using molecular identification such as DNA barcoding for medicinal plants could be very challenging at the same time in terms of generating barcode data and analyzing these data to stand on the discrimination power (Cowan and Fay 2012). A barcode is a short fragment sequence of a gene, which has been agreed upon and used to identify species based on the references contained in the BLAST or BOLD DNA sequence database (Kress et al., 2009). The selection of an accurate barcode is very challenging to distinguish very closely related or newly developed species. Species cannot be identified using DNA barcodes if the variation within the barcode between species is low or has a history of hybridization (Kress and Erickson 2008). Development of accurate DNA barcoding sequence is necessary for authentication and identification of plant species (CBOL 2009) The chloroplast rbcL genes (Ribulose -1.5 bisphosphate carboxylase/oxygenase large subunit), rpoC1 (RNA polymerase subunit C1) and nuclear region ITS (Internal transcribed spacer) consider as a most DNA barcoding genes used in the phylogeny of plants (Hollingsworth et al., 2016). Our study aimed to evaluate rbcL, rpoC1 and ITS DNA barcode sequences to

provide recommendations about the identification efficiency of *Clerodendrum phlomidis*.

2. MATERIAL AND METHODS

2.1.Taxon collection and authentication:

The taxon of C. phlomidis (syn. Volkameria multiflorum) was collected from the North Coast Region, Egypt. The species was identified and verified under the authority of the National Gene Bank (NGB), Agricultural Research Center (ARC), Giza, Egypt. Morphological identification was taken at the field during the collection mission, and herbarium voucher specimens at the NGB Herbarium, Taxonomy Dept., NGB. Observations followed the terminology by (El-Gazzar et al., 2019).

2.2.DNA extraction and purification:

The genomic DNA was extracted from 5g of fresh tissues using the CTAB-modified method (Elsherbeny, 2016). The extracted DNA was qualified and electrophoresed (5 μ l of DNA sample/100ml TE buffer) using 1.2% agarose gel and visualized by UV light. The extracted DNA was measured using the UV spectrophotometer (BIO-RAD- SmartSpec Plus spectrophotometer) between a range from 260nm to 280nm (Elsherbeny, 2016).

2.3.PCR and Amplification of Target Genes:

The PCR amplification used three universal genes viz., ITS, rbcL, and rpoC1 were chosen as presented in Table (1). The PCR reaction was performed with approximately 50 µl reaction mixture (1x Flexi buffer, 50ng DNA template, 2.5mM MgCl 2, 10uM dNTPs, 0.4uM of each primer, and 1U Pro mega© Green Go Taq TM enzyme). The PCR was optimized at 94°C for 5 min for an initial denaturation cycle, followed by 40 cycles of 94°C for 30 sec for the denaturation step, 50°C for 1 min for the annealing step, and 72°C for 30 sec for the elongation step, and 72°C for 7 min for the final extensions. The PCR product was run on 1.5% agarose gel, visualized under a trans illuminator UV light, photographed and recorded via the gel documentation system.

Marker	Primer and	d Length (bp)	
ITS	Forward	5`-ACGTCCCTGCCCTTTGTACACA-3`	(22 bp)
115	Reverse	5`-GCGGTACTTGTTGCTATCGGT-3`	(21 bp)
wh aT	Forward	5`ATGTCACCACAAACAGAGACTAAAGC-3`	(26 bp)
rbcL	Reverse	5`-GTAAAATCAAGTCCACCRCG-3`	(20 bp)
wa c1	Forward	5`- GGCAAAGAGGGAAGATTTGG -3`	(20 bp)
rpoC1	Reverse	5`- CCATAAGCATATCTTGAGTTGG -3`	(22 bp)
	F.	95°C 10 min	
		(94°C 30s, 55°C 30s,72°C 1min) x 35 cycles	
PCR conditions	R.	72°C 10min	

Table 1. Primer sequences and PCR conditions based on ITS, rbcL and rpoC1 barcodes are under study.

2.4.Sequencing of PCR products:

The Purified PCR products were sequenced according to the principles investigated by (Sanger *et al.*, 1977). Sequence Analysis Software recorded and collected sequenced data. The sequence data was performed with the same amplification DNA barcode primers.

2.5.Assignment of taxa:

BLAST software was applied to all recorded sequences of *Clerodendrum phlomidis* using NCBI online databases. The hits with maximal percent identity scores > 90% were recorded as successful when all involved a single genus.

2.6.Data analysis:

2.6.1. Genetic parameters based on type of barcode:

Nucleotide bases and their percentages with gap % based on ITS, rbcL and rpoC1 sequences of *Clerodendrum phlomidis* were estimated after sequencing.

2.6.2. Pairwise Alignment:

A pairwise alignment was carried out individually based on ITS, rbcL and rpoC1genes marker on NCBI database using BLASTN, some random sequences are selected, from BLASTN results ranging from 90 - 100% identity with the query sequence were performed to alignment on GenBank using ClustalW algorithm. The Sequence was assigned to taxon by comparing it with the sequences in the GenBank database. The E-value with lower percentage was more similar query sequence to the reference in the database. Reference sequences with their accession numbers were downloaded from GenBank (Janzen, 2019). The identified sequences have been submitted in GenBank.

2.6.3. Phylogenetic relationship examination:

Phylogenetic relationship clusters were reconstructed using NCBI with neighbor-joining software based on the aligned sequences. Based on the single locus alignment, Phylogenetic clusters were reconstructed (Tamura *et al.*, 2018).

2.6.4. Predicate amino acid of DNA sequence:

Translation and estimation of amino acid compositions based on ITS, rbcL, and rpoC1 nucleotide sequences carried out by computerassisted generated reading frames using online translation software (www.web.expasy.org/translate) according to (Gasteiger *et al.*, 2005).

3. RESULTS

3.1. Morphological identification:

Clerodendrum phlomidis were collected from the North Coast Region, Egypt. The plant material was morphologically identified, authenticated and data-passported under the authority of the National Gene Bank, Egypt, based on the rules of gene banks shown in Fig (1).



Figure 1. The herbarium leaves of *Clerodendrum phlomidis* for DNA isolation.

3.2.Taxa Identification:

The obtained data from ITS, rbcL and rpoC1 sequences of C. phlomidis revealed a total length of 714, 1968, and 509 bp, respectively. The amount of each base present within the nucleotide sequence with their respective percentages is presented in Table (2). The query of obtained three partial sequences was blasted at online GenBank databases individually. Over hundred reference sequences showed significant alignment with percent identity between 95 - 100%. The accessions KF199889 (98.7%) and ON010669.1 (98.6%) were revealed by ITS and rbcL sequences identical to the query taxon of C. phlomidis, as shown in Table (3&4). On the other hand, the query sequence revealed similar to the taxa of C. buchneri, C. cephalanthum, C. thomsoniae, and C. rotundifoliuma by ITS sequences Table (3), C. indicum, C. thomsoniae, C. longiflorum, C. infortunatum, C. floribundum, C. cephalanthum, C. buchneri, and C. splendens by rbcL Table (4) C. japonicum, C. thomsoniae, C. cyrtophyllum and C. trichotomum by rpoC1 sequences Table (5). These taxa have a high identity to the query of C. phlomoids which range from 93% to 98% in ITS, from 98% to 99% in rbcL, and from 98.7 to 98.9% in rpoC1. In the case of the ITS sequence (714 bp), the closest species was C. phlomidis with an identity percentage of 98.74%. In the rbcL sequence (1968 bp), the closest species was C. indicum with an identity percentage of 98.89%. Finally, in the rpoC1 sequence (509 bp), the closest species was C. japonicum with an identity percentage of 98.96%.

 Table 2. The percentages of nucleotide bases of C. phlomidis based on sequenced ITS, rbcL and rpoC1.

NO.	Barcode Name	Seq.	_	Ba	ses		Gap %	GC	GC %
	Darcoue Name	Length	Α	Т	С	G			
1	ITS	714	166	140	200	208	50.35%	408	57.14%
2	rbcL	1968	516	359	370	723	5.84%	1093	55.54%
3	rpoC1	509	132	161	99	117	70.56%	216	42.44%

No.	Scientific Name	Length (bp)	Query Cover %	E-value	Identity %	Accession
1	Clerodendrum phlomidis	655	88%	0.0	98.74%	KF199889
2	Clerodendrum buchneri	674	92%	0.0	95.65%	KT728417
3	Clerodendrum cephalanthum	686	90%	0.0	95.83%	JN575348
4	Clerodendrum rotundifolium	691	92%	0.0	93.24%	U77766
5	Clerodendrum thomsonae	697	92%	0.0	93.40%	U77745
6	Clerodendrum thomsonae	689	92%	0.0	93.24%	U77743
7	Clerodendrum thomsonae	596	83%	0.0	96.48%	AF477778
8	Clerodendrum thomsonae	492	55%	7e-172	94.95%	KC896701
9	Clerodendrum thomsonae	485	53%	3e-165	94.56%	KC896700

 Table 3. Description of the Identity percentage between C. *phlomidis* and the queries based on the partial ITS region against NCBI database.

 Table 4. Description of the Identity percentage between C. phlomidis and the queries based on the rbcL gene against NCBI database.

	Total gene agamst 1(ODI data					
No.	Scientific Name	Length (bp)	Query Cover %	E-value	Identity %	Accession
1	Clerodendrum indicum	694	27%	0.0	98.89%	MK331785.1
2	Clerodendrum phlomidis	562	26%	0.0	98.62%	ON010669.1
3	Clerodendrum thomsoniae	706	27%	0.0	98.71%	MK331803.1
4	Clerodendrum longiflorum	573	27%	0.0	98.71%	KU564780.1
5	Clerodendrum infortunatum	536	27%	0.0	98.69%	JQ724863.1
6	Clerodendrum infortunatum	536	27%	0.0	98.69%	JQ724864.1
7	Clerodendrum floribundum	566	27%	0.0	98.87%	KF496552.1
8	Clerodendrum cephalanthum	552	26%	0.0	99.05%	KU568049.1
9	Clerodendrum buchneri	552	27%	0.0	98.89%	KU568050.1
10	Clerodendrum splendens	552	27%	0.0	98.89%	KX783849.1

 Table 5. Description of the Identity percentage between C. phlomidis and the queries based on the rpoC1 gene against NCBI database.

No	Scientific Name	Length (bp)	Query Cover %	E-value	Identity %	Accession
1	Clerodendrum japonicum	152,217 C. DNA	94%	0.0	98.96%	MW181770.1
2	Clerodendrum japonicum	152,279 C. DNA	94%	0.0	98.96%	MT473745.1
3	Clerodendrum japonicum	152,215 C. DNA	94%	0.0	98.96%	MW307827.1
4	Clerodendrum japonicum	152,171 C. DNA	94%	0.0	98.96%	NC_056260.1
5	Clerodendrum thomsoniae	151,053 C. DNA	94%	0.0	98.75%	OM912812.1
6	Clerodendrum thomsoniae	151,053 C. DNA	94%	0.0	98.75%	NC_064126.1
7	Clerodendrum thomsoniae	151,053 C. DNA	94%	0.0	98.75%	OM617840.1
8	Clerodendrum cyrtophyllum	152,004 C. DNA	94%	0.0	98.75%	MW858153.1
9	Clerodendrum cyrtophyllum	152,004 C. DNA	94%	0.0	98.75%	MW858153.1
10	Clerodendrum trichotomum	151,693 C. DNA	94%	0.0	98.75%	MT473746.1

3.3.Phylogenetic analysis:

The molecular phylogenetic trees are represented in a linear form using neighborjoining methods based on the BLAST-NCBI Genbank database. The phylogenetic clusters was reconstructed for each sequence based on closest species with the estimated substitution matrix shown in Figures (2,3&4). In the ITS sequences, the species divided into two main groups. The first group (left cluster) includes *C. rotundifolium* only; while, the second group (right cluster) is divided into more clusters. It is notably that the taxa of *C. buchneri* and *C. cephalanthum* fall into a cluster Fig (2). In the rbcL sequences, the species divided into two main groups. The first (left cluster) includes group С. phlomoidis (ON010669.1) and our taxa (C. phlomoidis) under study, while, the second group (right cluster) is divided into more clusters, one of these clusters contain more than 5 accessions of Volkameria inermis, this species is another name of C. phlomoidis Fig (3). In the rpoC1 sequences, the species divided into two main groups. The first group (left cluster) includes only our taxa (C. phlomoidis) under study, while, the second group (right cluster) is divided into more clusters, three accessions named C. thomsoniae fall into a cluster and four accessions of C. japonicum fall into another cluster Fig (4).

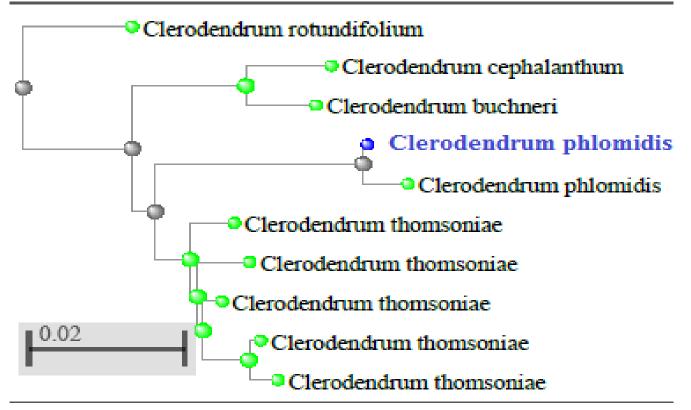


Figure 2. Distance tree of results between the queries and *C. phlomidis* based on the partial ITS region against NCBI database.

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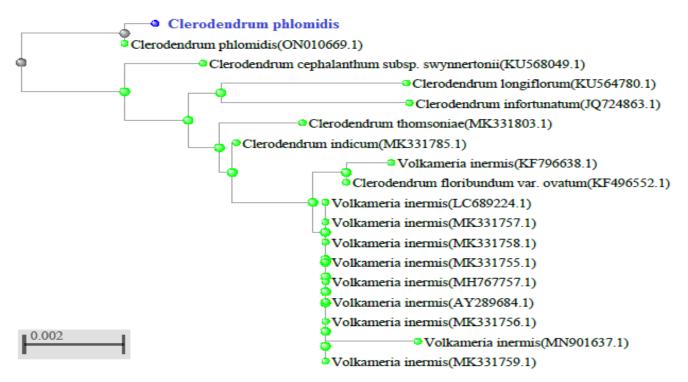


Figure 3. Distance tree of results between the queries and *C. phlomidis* based on the rbcL gene against NCBI database.

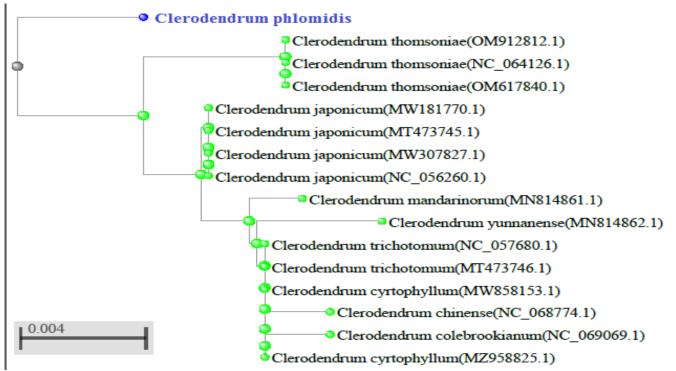


Figure 4. Distance tree of results between the queries and *C. phlomidis* based on the rpoC1 gene against NCBI database.

3.4.Sequence Alignment:

The alignment is done with the closest taxa derived by three genes viz., ITS, rpoc1and rbcL sequences. The ITS sequence revealed the identical taxon to *C. phlomoidis* (KF199889.1) with a similarity of 98.74%; where the difference in alignment scored 8 nucleotides with a gap of 4 nucleotides, as shown in **Fig** (5). In the rbcL results, the closest taxon recorded with *Clerodendrum indicum* (MK331785.1) with a similarity of 98.89%. The variance scored 5

Sequence ID: KF199889.1 Length: 655 Number of Matches: 1

nucleotides with a gap of 2 nucleotides Fig (6). On other hand. the closest the species was Clerodendrum japonicum (MW181770.1) with a similarity of 98.96%. The genetic variation recorded 5 nucleotides without gap, as presented in Fig (7). Sequence alignment (SA) is a method for detection variations and similarities between DNA nucleotide sequences. It is important analysis in bioinformatics for characterize functional or structural relation between sequences and is effective for biodiversity analysis (Zutau, 2017).

Clerodendrum phlomidis isolate FRLH-L/10/11/10 18S ribosomal RNA gene, partial sequence

					indicineo.			
_	1:19	to 655 Ger						
Score 1129 b	hits(61	1)	Expect 0.0	Identities 629/637(99%)		Gaps 4/637(0%)	Strand Plus/Pl	
Query Sbjct	10 19			TCG-AACCTGCATA				65 78
Query Sbjct	66 79			GCGGTCCCCTCATC				125 138
Query Sbjct	126 139			GCGGAATGCGCCAA				185 198
Query Sbjct	186 199			ATCGTGGGGGAGGTT				245 258
Query Sbjct	246 259			GGCTCTCGCATCGA				305 318
Query Sbjct	306 319			CGTGAACCATCGAG				365 378
Query Sbjct	366 379			CTGCCTGGGCGTCA				425 438
Query Sbjct	426 439			GATATTGGCCTCCC				485 498
Query Sbjct	486 499			GAAAGTCACGACCA				545 558
Query Sbjct	546 559			CGTCCGATCGGGAG				605 618
Query Sbjct	606 619			GACCCCAGGTCAGG		642 655		

Figure 5. Alignment between the highest identities query and *C. phlomidis* based on partial ITS region against NCBI database.

Clerodendrum indicum isolate Cind1 ribulose-1,5-bisphosphate carboxylase/oxygenase large subunit (rbcL) gene Sequence ID: <u>MK331785.1</u> Length: 694 Number of Matches: 1

Score 965 bit	s(522) Expect	Identities 535/541(99%)	Gaps 2/541(0%)	Strand Plus/Plu	us
Query Sbjct	32 23		TTTATTATACTCCTGAATA			91 80
Query	92 81		CTCCTCAACCTGGAGTTCC			151 140
Query	152 141		CTGGTACATGGACAACTG			211
Query	212 201		GATGCTACCACATCGAGG			271
Query	272 261		ATCCTTTAGATCTTTTGA			331 320
Query	332 321		ATGTATTTGGATTCAAAGO			391 380
Query	392 381		CTTATATTAAAACTTTCC			451 440
Query	452 441					511 500
Query	512 501		AAAACTATGGTAGAGCGG			571 560
Query	572 561	A 572 . 561				

Figure 6. Alignment between the highest identities query and *C. phlomidis* based on rbcL gene against NCBI database.

Clerodendrum japonicum chloroplast, complete genome Sequence ID: <u>MW181770.1</u> Length: 152217 Number of Matches: 1

Range 1: 21435 to 21914 GenBank Graphics

Score 859 bit	ts(465)	Expect 0.0	Identities 475/480(99%)	Gaps 0/480(0%)	Strand Plus/Minus	
Query Sbjct	18 21914		GACGTTCCGTCATTGTCG			77 21855
Query Sbjct	78 21854		AAATAGCAATAGAGCTTTI			137 21795
Query Sbjct	138 21794		CTTCGAACATAGGAGTTGC			197 21739
Query Sbjct	198 21734		TACTGCAGGAAGTTATGCA			257 21679
Query Sbjct	258 21674		AATTGGGCATACAGGCATT			317 21619
Query Sbjct	318 21614		CATTAGTTTGTAAGGGATT			377 21555
Query Sbjct	378 21554		CCTTATCTTTGGAGGCTCA			437 21499
Query Sbjct	438 21494		TGTCTCCAGCTATTGGAGA			497 21439

Figure 7. Alignment between the highest identities query and *C. phlomidis* based on rpoC1gene against NCBI database.

3.5.In silico translation of DNA sequence:

In silico translation of the ITS, rbcL, and rpoC1 barcode nucleotide sequences carried out into three reading frames (Protein sequences). Figures (8,9&10) displays Frames 1 - 3 (5'-3'). Nucleotide sequences in DNA or RNA can be divided into a collection of sequential, non-overlapping triplets called reading frames (RFs), which translated to stop codons or amino acids (Pearson *et al.*, 1997). DNA sequences designated as open reading frames (ORFs) in the genome tend to be translated into amino acids (Yazhini, 2018). Previously,

only large ORFs (> 300 nucleotides) were used to be protein-coding areas, which are regarded to be genes, but the discovery of short ORFs (less than 100 nucleotides) led to discovered many translating functional small ORFs and presence of their stable peptide chain. As a result, we checked the small ORFs contained within our sequences, given that some exons are extremely short. Consequently, Finding ORFs in higher eukaryotic sequences is significantly genome more challenging than in prokaryotic genomes, despite of the exons of protein-encoding genes must be ORFs (Parker, 2001).

5'3' Frame 1	
S L V L R R H C R T C I A D R E H V F K Q I G A A	VFCGPLIAGVRQRVAVRSN
KIGRG Met R Q G I H K R V F P S P G P V C G D) R G E V G Met P V V Y K N D S R Q R I
S R L S H R Stop R T Stop R N A I L G V N C R I P	stop T I E S L N A S C A R S H Stop A
EGTSAWASRITSPPSTHSAVDGGGY	. W P P V H H S C A A G P N A I P R W R
K S R P V L V E V S T R V L S Stop H K T S S D R	ESLQTQWRIHALRLRPRPQ
VRRDYPLSLSIS Stop PGKKKTCVKSF	RRRG
- 5'3' Frame 2	
P Stop Y C G D I V E P A Stop Q T A N T C L N K S	GLESSAVPSSPACANASTC
G L T K S G A E C A K E Y T K E C S P P O G P C A	
YLGSRIDEERSE Met RYLV Stop IAES	
ARLPGRHASRRLPPHTVLLMetGADI	-
ESHDQCWLKYQLACCRDTRRPIGS	HYRPSGAFTHCASDRDPRS
GGITR Stop V Stop A YHNLERKKPA Stop K	KVEGGG
5'3' Frame 3	
L S T A E T L S N L H S R P R T R V Stop T N R G	
Stop Q NRARNAPRNTQKSVPLPRARV	
DISALAS Met KNVAKCDTWCELQNPV	
C L G V T H H V A S L H T Q C C Stop W G R I L A V T T S V G Stop S I N S R A V V T Q D V V R S G	
QAGLPAEFKHIITWKEKNLREK Stop K	
QAGLEALERALITWKERNLKER Stop	A E A G

Figure 8. ITS nucleotide sequence of *C. phlomidis* translated to 3 reading frames using Expasy online translation software.

5'3' Frame 1
T Mat 5 Å R Q R G À L Stop V T N Stop L Y Y T P E Y E T K D T D I L À À F R V T P Q P G V P P E E À G À À V À À E S S T G T M T T V M T D G L T S L D R Y K G R C Y H I E À
V L G E K D Q Y I C Y V A Y P L D L F E E G S V T N Ket F T S I V G N V F G F K A L R A L R L E D L R I P T A Y I K T F Q G P F H G I Q V E R D K L N K Y G R P L L G C T I
X P K L G L S A K N Y G R A V Y E C L R G G L E V F L K K R T Y T Met I L T K Stop Stop H T C Y S A R E Q S V S S V L D Y E Y F Met T G G G G E E R K H H E E I Y D E E G V
R Mat V T I D K E T I T V R R E S D D R G À A G G E E R G G À D Stop N G N R G G À E E E R R L E K R G D V D D I G T P À V R V V M G D À S G À D Q D R E C D G G M R Stop
E T G W D I R G E D G G R V G G R Met C T R R Stop V K G R Q T D E A E G V R I W V G V G N R G R G G Stop E N G G E R S G S E A R G T Stop R Q E R F G E P C G I C E G G V D
A Q A A G G A G R Q G R H S H R T E R T K D E Stop G G G R V D A G C E T E S R G G T T G E R P G T H T A R G T G R A N R Mat H V V N S R R C I V Stop L R Q R R N R S Stop D
K Stop S Q S R I W S G A S I E G D Stop S A Stop R K T R Y D I Stop R D E W L S T G R P D D R Met G S Met R D G Met E P Q I A G L R T D A G X R A R H R Met V E A D R A A R
D G & R Q R L R M C G R Stop D & R & Stop R C Q R E Y E E T V G E R R P Stop E T C & N T E Mat R P T T D E R D & G M G X R Stop T S D D M E G Q T S R E Mat C Q V I N C V
TRERCTWAGE
- SY Frama 2
53 Frame 2
Q C L Q D S E E H Y E L Q I D F I I L L N T K P K I L I S W Q H S E Stop L L N L E F R L K K Q G L R Stop L P N L L L V H G Q L C G P Mot A L F A L I V T K V D A T T S R F
F L E K K I N I F V Met Stop L I L Stop I F L K K V L L L T C L L P L Stop E Met Y L D S K P Y V L Y V W K I C E S P L L I L K L S K A R L Met G S K L K E I N Stop T S T V
V L C W D V L L N R N W G Y P L K T Met V E R F Met N V F A V D L R F F Stop K N E H T L Stop F S Q S D D T R A I P R E S R A S Q V Y W T T N I L Stop Q E G G A K S A S T
Mat RRS Mat RAE Stop IN LRStop TRRLStop P Stop I GRATIGG PRVERSEVGGRIR Mat EIGAGRRKRGD N RREETS Mat TLGRRRSGSCGG
TRAGRTRIGSATEGGDERRAGTYAARTGGE Stop AGGCARGVRSKDDRRTRRRACGYGWGSGIGAGGGER Met GVNGADRRRVGRDGKS
G R A S R A G Y A K V G N Mat H R O R E V L G G K E D T H I G P R G R R T S E V G G A M T O D A R O N R E A G R O A S G R G R I P R V E R V A R I E C T S Stop T V A G A
Stop S S Y G R D V I D R K I S D H R A A S G O E R R L R E T E A R S G R P D Wat I Y D A Mat S G Stop A L V G R T I G W A A Stop E Wat V W N H K S R D C E R Mat L E S E L V
I E W W K P I V Q R E Met E R G S D C D G V A D E T R E R R G V N G S Met R K Q W E S V D R E R R V R T P R C G R R R T N A Met Q G D G G S A E R R Met T G K G R H R A R C
V K Stop L I V S R G N D V H G L G
5'S' Frame S
N V C K T A R S T Mat S Y K L T L L Y S Stop I R N Q R Y Stop Y L G S I P S N S S T N S S A Stop R S R G C G S C R I F Y N Y Mat D N C V D R N P Y Q P Stop S L Q R S Mat L
FRAGRSWRKRSIYLLCSLSFRSFIStopRRFCYStopHVYFHCRKCIWIGSPTCSTSGRSANPHCLYStopNFPRPASWDPSStopKRStopIEG
VRSSSVG Met YY Stop TEIGVIR Stop KLW Stop SGL Stop Met SSRWT Stop GFFKKTNIHYDSHKV Met THVLFRERAERLKCIGLRIFYDRRGG
R R A Q A P Stop G D L Stop Stop G G S E N G Y D R Q G D Y N R E K G E R R Stop G G R G N R G A R N V G G L E N K Stop G R G G R E A T G E E R R R S top H N D A G F G
R V G G R E R G G F G Stop G V R R R V A Met R D G L G H T R R G R G A S R R E D V H E A L G Q R T T D G R G G G R A D Met G G G R E S G Q G G V R E M G Stop T E R I G G A
N D V T A R A A G R A V R D Met R R M G G C T G S G R C M A A R K T L T S D R E D E G R V R M G A R G R R Met R D R I A R R D D R R A A G D A Y R A M N G S R E Stop N A R R
E Q S F V H S L A T A E T Stop S I V R Stop V I T E F H L V R S V D Stop G R L K R V A E D Q I Stop Y Met T R Stop V A E H W S A G R Stop D G Q H E R W Y G T T N R G T A N
G C W K A S S S S N G G S R S C S E R W S E A A I A Met V W P Met R E S V E V S T G V Stop G N S G R A S T V R D V C E H R D A A D D G R T R C R G Met G E A L N V G Stop
L G R A D I A R D V S S D Stop L C H A G T Met Y Met G N A

Figure 9. rbcL nucleotide sequence of *C. phlomidis* translated to 3 reading frames using Expasy online translation software.

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S	C	I	P	E	Ξ	st	or	>	s	т	н	s	A	s	te	pp	I	G	н	т	G	I	F	A	H	E	- 5	G	G	A	C	Y	I	5 1	F 1	г	s	I	s	L	st	op	G	I	\$
C	R	F	s	te	pp	W	0		s	N	G	C	S	C	: 1	r 1	6	I	F	G	G	s	s	G	G	P	F	т	Y	v	F	s	Y	E	s	F	v	s	S	Y	W	R	s	н	1
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3	I	S	to	p	н	s	to	pp	S	З	C]	E (0	D	v	P	S	L	S	s	to	P	v	L	н	F	н	Y	I	D	v	D	Y	R	v	K	s	to	P	Q	st	op	S	F	1
2	н	L	s	to	pp	F	V	7 7	v	st	op	, 1	G. 1	D	N	I	L	L	R	т	St	tor	> 1	E I	L 1	L I	R	7 1	< 1	F G	F	C I	K I	N	st	op	L	Y	G	K	Y	C	R	K	1
2	R	G	I	1	5	Y	C	s	to	P	I	E	H	F	, 1	6 0	2	Ι	N	W	A	Y	R	H	S	S	P	F	st	op	W	F	2 0	3 1	v i	L	F	v .	Y	I	н	st	op	F	3
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Figure 10. rpoC1 nucleotide sequence of *C. phlomidis* translated to 3 reading frames using Expasy online translation software.

4. **DISCUSSION**

Plant DNA barcoding works best when coding and noncoding genetic markers are combined. The plastid-conserved rbcL gene and the more variable rpoC1 gene are the most commonly investigated markers in several studies. Recently, the ITS region has also shown itself to be an effective variable marker (Kress, 2017). Therefore, the purpose of this study was to determine for the first time how efficiently these markers worked as DNA barcoding for the rare Egyptian plant Clerodendrum phlomidis. Except for the pairs used for the rbcL gene, which may require specificity for successful more amplification, the primers used in the amplification of the examined markers under study were sufficient, although it is not particularly useful in identifying plant species on their own, we have other markers to corroborate the identification of plant genus, so this is not much of a barrier in this case (Kang et al., 2017). The primers utilized for amplification in each case were also used to partially sequence the remaining markers, which included the ITS region, rbcL and rpoC1 genes. The identification of the plant species C. phlomidis has been confirmed by the results. The ITS region, rbcL and rpoC1 genes have been demonstrated in numerous studies to be highly effective genetic markers for identifying Clerodendrum species, making them promising candidates for plant DNA barcoding (Han et al., 2016)(Yu et al., 2021). Numerous Clerodendrum species were discovered to be comparable to the rbcL rpoC1gene sequences. and which

demonstrated efficacy in identifying the species when both pairs of primers were used, although the closest was C. indicum and C. japonicum respectively. Using phylogenetic analysis, one may determine potential evolutionary paths for the nucleotide family. By representing the sequences as outer branches of a tree, the evolutionary ties between them are illustrated. The degree to which various sequences are related is then reflected in the branching relationships on the inner portion of the tree. (Mount, 2001). If two DNA molecules from different organisms have identical sequences, it is likely that previous generations inherited these DNA sequences (Erickson et al., 2008). Egyptian Clerodendrum phlomidis was identified and documented here in this study with three DNA barcode sequence for the first time. The taxon was matched to neighboring taxa in each barcode sequence and the phylogenetic clusters were reconstructed. Additionally, by aligning between C. phlomidis and C. indicum sequences based on rbcL gene and another aligning between C. phlomidis and C. japonicum sequences based on rpoC1 gene obtained a single sequence with high identity, the identification process produced identical findings, sequence's supporting the utility as а differentiating marker of the C. phlomidis plant. Regarding the ITS region, differences in the relatedness of *Clerodendrum* species to *C*. phlomidis were found in the sequences produced using the ITS primers and their alignment. This provided yet another argument in favor of the ability of species to discriminate in this area. The two distinct sets of primers allowed for the sequencing of the ITS region. successful Typically, even with good amplification, sequencing the ITS region can provide some challenges (Yu et al., 2021)(Wang et al., 2016).

5. CONCLUSION

It can be said that the usefulness of these partial sequences viz., ITS, rbcL and rpoC1 genes are reliable markers for *C. phlomidis* DNA barcoding and identification, especially two genes of rbcL and ITS because of their linkage to *C. phlomoids*. Even if the result of rpoC1 was not particularly encouraging, more specifically designed primers can produce better results. Researchers can use a standard DNA region as a DNA barcoding to identify unknown species. The results of this research proved that it is possible to ITS, rbcL, and rpoC1 genes in *C. phlomidis* successfully amplified and sequenced. Furthermore, the ITS region, rbcL and rpoC1 gene sequences have a 98–100% identity to taxa from the Lamiaceae family, according to the results of BLASTN analysis, single sequence alignment, and phylogenetic analysis.

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الملخص العربي

التعريف الجيني و شجرة القرابة الوراثية لنبات Clerodendrum phlomidis في مصر باستخدام تقنية ترميز الحمض النووي (الباركود)

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